

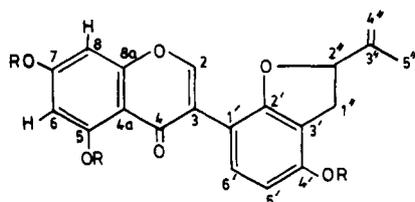
ISOFLAVANOIDS OF *CROTALARIA MADURENSIS*¹

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In continuation of our earlier studies (1,2) on the chemistry of the stem and leaves of *Crotalaria madurensis* R. Wight (Leguminosae), we now report the isolation and structure elucidation of two new isoflavones named crotarin [**1**] and crotalarin [**3**]. These isoflavones were obtained by repeated silica gel chromatography of the CHCl₃-soluble fraction of the EtOH extract followed by purification through preparative tlc.

The molecular formula of crotarin [**1**], C₂₀H₁₆O₆, was determined by ms (M⁺, *m/z* 352). Its ir spectrum showed strong bands at 3225, 1660, and 1640 cm⁻¹ suggesting the presence of an hydroxyl group and an α , β -unsaturated keto moiety. The uv absorption maximum at 262 and 324 (sh) nm of **1**

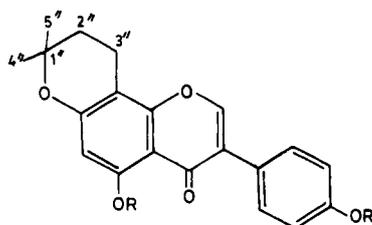


1 R=H
2 R=Ac

the aromatic region, a singlet at δ 7.8 (1H) was characteristic of the C-2 proton (5) of an isoflavone nucleus, while a three-proton multiplet at δ 6.2 corresponded to aromatic protons at C-6, C-8, and C-5'. The *ortho* coupled doublet at δ 6.84 (*J*=12 Hz) was due to the C-6' proton.

The retro Diels-Alder fragmentation of **1** (M⁺, *m/z* 352) in eims gave an ion at *m/z* 200 (C₁₃H₁₂O₂) comprised of ring B, while the ion constituting ring A appeared at *m/z* 153. The ion of *m/z* 200 further showed the loss of a methyl group to give the fragment at *m/z* 185.

The ¹H-nmr pattern coupled with the mass fragmentation confirmed the presence of a dihydrofuran ring having an isopropenyl group in ring B of crotarin.



3 R=H
4 R=Ac

and shifts with AlCl₃ and NaOMe indicated the presence of an isoflavone nucleus (3). The ¹H-nmr spectrum of **1** revealed the presence of a dihydrofuran ring having an isopropenyl group (4). Methyl protons of the isopropenyl group at C-5'' appeared as a singlet at δ 1.70. Two singlets integrating for one proton each at δ 4.8 and 4.98 were assigned to the exocyclic methylene protons at C-4''. A triplet at δ 5.1 integrating for one proton accounted for the C-2'' proton. In

Crotarin [**1**] formed a triacetate **2** (M⁺, *m/z* 478). The ¹H-nmr spectrum of **2** exhibited three peaks for acetoxy methyls at δ 2.08, 2.24, and 2.30, confirming the presence of three hydroxyl groups. In the aromatic region, the position of *ortho*-coupled proton of ring B at C-6' remained unaffected, while the C-5' proton showed a paramagnetic shift to δ 6.9 and appeared as a doublet. The *meta*-coupled C-6 and C-8 protons shifted downfield to δ 6.78 and 7.14, respectively, inferring the *ortho*-relationship of these protons to the site of acetylation. Thus, the three hydroxyl groups

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in crotarin were placed at C-5, C-7, and C-4'.

The structure of crotarin as **1** was further supported by ^{13}C -nmr spectral studies (Table 1). The broad band de-

TABLE 1. ^{13}C nmr of Crotarin [**1**] and Crotaralin [**3**] in $\text{DMSO}-d_6$, TMS as Internal Standard

| Carbon | Compounds | |
|-----------------|-----------|-----------|
| | 1 | 3 |
| C-2 | 155.16, d | 151.1, d |
| C-3 | 120.60, s | 124.0, s |
| C-4 | 180.46, s | 174.5, s |
| C-4a | 104.48 | 106.4, s |
| C-5 | 161.17, s | 158.1, s |
| C-6 | 98.79, d | 94.19, d |
| C-7 | 164.06, s | 157.7, s |
| C-8 | 93.53, d | 109.12, s |
| C-8a | 157.60, s | 155.9, s |
| C-1' | 112.6, s | 125.5, s |
| C-2' | 152.08, s | 131.1, d |
| C-3' | 111.32, s | 115.7, d |
| C-4' | 160.84, s | 160.7, s |
| C-5' | 100.16, d | 115.7, d |
| C-6' | 131.76, d | 131.1, d |
| C-1'' | 31.97, t | 75.74, s |
| C-2'' | 85.28, d | 31.74, t |
| C-3'' | 143.94, s | 17.66, t |
| C-4'' | 111.01, t | 27.30, q |
| C-5'' | 16.91, q | 27.30, q |

coupled spectrum of the compound exhibited signals for 20 carbon atoms, and the SFORD spectrum revealed the presence of 11 quaternary, 6 methine, 2 methylene, and 1 methyl carbons. The signal at δ 180.4 was assigned to the carbonyl carbon (C-4) of the isoflavone nucleus (6). The characteristic C-2 carbon (7) of **1** appeared as a doublet at δ 155.2. The five oxygen bearing carbons in the aromatic ring (8) appeared between δ 164.06 and 152.08. The downfield carbons at δ 164.16 and 161.17 could be assigned to the C-7 and C-5 carbons, while C-8a appeared at δ 157.6 (9). The chemical shifts of the oxygen bearing aromatic carbons were in agreement with the *meta*-substitution rather than occupying *ortho*-positions. The chemical shifts for *ortho*-substituted oxygen-bearing carbons are reported (10) to be be-

tween δ 144.0 to δ 149.0, and, hence, isoprenyl group in ring B, constituting the dihydrofuran ring, is attached through C-3' to oxygen at C-2'. The alternate cyclization of the isoprenyl group attached at C-2' to oxygen at C-3' is ruled out as the latter would place the C-3' and C-4' oxygens *ortho* to each other and would have influenced the ^{13}C chemical shifts of C-3' and C-4' (10). The doublet at δ 85.28 was assigned to C-2'' and the triplet at δ 31.97 to C-1'' of the dihydrofuran ring (11). The quaternary carbon of the isopropenyl group appeared at δ 143.94 and the methyl at δ 16.91, while the methylene carbon was observed as a triplet at δ 111.01. Due to the shielding effect of the *ortho*-substituted hydroxyl function, C-6 and C-8 in ring A and C-5' in ring B appeared at δ 98.79, 93.53, and 100.16, respectively. This fully supported the structure of crotarin as **1**.

Crotalarin [**3**] was obtained as colorless needles, mp 281-85°, and the molecular formula $\text{C}_{20}\text{H}_{18}\text{O}_5$ was confirmed by ms (M^+ , m/z 338). The ir spectrum of **3** has absorption bands at 3200 (OH), 1620, and 1560 cm^{-1} characteristic of an isoflavone carbonyl. The uv absorption at λ max 262 and 320 (sh) nm was supportive of the isoflavone structure (3) for crotaralin **3**. The ^1H -nmr spectrum of **3** suggested the presence of a dimethyl chroman ring (12,13). The methyl protons of the dimethyl chroman ring at C-4'' and C-5'' appeared as a singlet at δ 1.3, and two multiplets at δ 1.74 and 2.6, each integrating for two protons accounted for the methylene protons at C-2'' and C-3'', respectively. A characteristic singlet of the C-2 proton of the isoflavone nucleus in **3** appeared at δ 7.62. The pair of *ortho* coupled C-2', C-3' and C-5', C-6' protons appeared at δ 6.75 and 7.32, respectively, while a singlet at δ 6.36 was assigned to C-6 proton indicating the C-alkylation at C-8.

The mass spectrum of crotaralin [**3**] showed the molecular ion peak M^+ at

m/z 338. The other significant peaks at m/z 323 ($M^+ - CH_3$), 295 ($M^+ - 43$), 283 ($M^+ - 55$), 282 ($M^+ - 56$) were characteristic of fragmentation of the dimethyl chroman ring (14). The peak at m/z 118 supported the presence of the monohydroxy (4'-OH) ring B.

Crotalarin [3] formed a diacetate 4 (M^+ , m/z 422). The 1H -nmr spectrum of the acetate 4 showed the presence of two acetoxy methyls at δ 2.22 and 2.3. The C-6 protons were shifted downfield by 0.3 ppm to δ 6.66 indicating its *ortho*-relationship to the site of acetylation (C-5). The C-3' and C-5' protons of ring B were also shifted downfield to δ 7.05 confirming the position of the hydroxyl group at C-4' in 3.

The structure 3 of crotalarin was further supported by the ^{13}C -nmr spectrum (Table 1). The C-4 carbonyl in 3 appeared at δ 174.5. The characteristic doublet at δ 151.1 accounted for the C-2 carbon of the isoflavone nucleus in [3]. The carbons bearing hydroxyl groups appeared at δ 160.7 (C-4') and 158.1 (C-5). Carbons of the dimethyl chroman ring resonated at δ 75.74 (C-1''), 31.74 (C-2''), 17.66 (C-3'') and two methyl carbons at δ 27.30 (C-4''), C-5'') (14). The doublet at δ 94.19 was attributed to C-6 experiencing an α -inductive effect due to the oxygen functions at C-5 and C-7 (10). Singlets at δ 157.7 and 109.12 were assigned to the C-7 and C-8 carbons, respectively. The carbons of ring B were centered at δ 125.5 (C-1'), 131.1 (C-2', 6'), 115.7 (C-3', 5'), and 160.7 (C-4') supporting the position of the C-4' hydroxyl group in crotalarin 3. Neither 1 nor 3 showed any antifungal activity.

EXPERIMENTAL

GENERAL EXPERIMENTAL PROCEDURES.—

Unless otherwise stated, uv spectra were obtained in MeOH, ir spectra were run KBr discs, and nmr spectra were obtained in $CDCl_3$ or a mixture of $CDCl_3$ and $DMSO-d_6$ with a Varian F-90 spectrometer. Tlc was carried out on Si GF 254 and column chromatography over Si gel (BDH).

PLANT MATERIAL.—The leaves and stems of

C. madurensis were collected in November-December 1978, from Kurnool, Andhra Pradesh, India (1), and a voucher specimen No. 110601 of the same is preserved in the herbarium of the Institute.

EXTRACTION.—The air dried plant material (5 kg) was pulverized and extracted with 95% EtOH (3×4 liters) at room temperature. The solvent from the percolate was removed under reduced pressure below 40°, and the resulting residue was fractionated as reported earlier (1) to yield the following fractions, petroleum ether (50 g), $CHCl_3$ (40 g), and EtOAc (30 g) from the non-alkaloidal part of the EtOH extract.

CHROMATOGRAPHY OF $CHCl_3$ -SOLUBLE FRACTION.—The $CHCl_3$ -soluble fraction (40 g) was chromatographed on a column of Si gel (1.5 kg). The column was successively eluted with C_6H_6 , C_6H_6 -MeOH (99:1), C_6H_6 -MeOH (98:2), C_6H_6 -MeOH (95:5), and C_6H_6 -MeOH (90:10), and 75 fractions (500 ml each) were collected.

CROTARIN [1].—Elution of column with C_6H_6 -MeOH (99:1) (fractions 21-35) gave a mixture which was further resolved by preparative tlc using C_6H_6 -MeOH (96:4) as eluent. The major compound thus obtained was crystallized from MeOH to give crotarin [1] (70 mg); (mp 260° dec); yield 0.0014%; λ max (MeOH) 262 nm (log ϵ , 5.4687), 324 (sh) (log ϵ , 4.1236) nm; λ max (MeOH)+ $AlCl_3$ 270 and 360 (sh) nm; λ max (MeOH)+ $NaOMe$ 262 and 324 nm; ir ν max (KBr) 3225, 1660, 1640, 1620, 1560, 1500, 1460, 1365, 1300, 1260, 1180, 1100, 1050, 975, 918, 825, 800, 740 cm^{-1} ; 1H nmr δ ($CDCl_3$ + $DMSO-d_6$) 1.70 (s, 3H, C-5''- CH_3), 2.7-3.5 (m, 2H, C-1''- H_2), 4.8 (s, 1H, C-4''-H), 4.98 (s, 1H, C-4''-H), 5.12 (t, $J=7$ Hz, 1H, C-2''-H), 6.14 (d, $J=2$ Hz, 1H, C-6-H), 6.26 (d, $J=2$ Hz, 1H, C-8-H), 6.28 (d, $J=7$ Hz, 1H, C-5'-H), 6.84 (d, $J=9$ Hz, 1H, C-6'-H), 7.8 (s, 1H, C-2-H); ^{13}C nmr (Table 1); eims m/z (rel. int.) 352 (M^+ , 52), 337 (M^+ , 52), ($M^+ - 15$, 100), 200 (8), 187 (22), 185 (12), 153 (12).

CROTARIN TRIACETATE [2].—A mixture of 1 (20 mg), Ac_2O (0.5 ml), and pyridine (0.5 ml) was kept at ambient temperature for 15 h. The resulting mixture was poured over crushed ice. The precipitate was washed with H_2O to neutral pH to give the triacetate 2 (18 mg), mp 196° (MeOH); ir ν max (KBr) 1760, 1620, 1590, 1270, 1200, 1160, 1135, 1045 cm^{-1} ; 1H nmr δ ($CDCl_3$) 1.72 (s, 3H, C-5''- CH_3), 2.08, 2.24, 2.3 (s each, 3H each, 3×OAc), 2.7-3.4 (m, 2H, C-1''- H_2), 4.82 (bs, 1H, C-4''-H), 5.0 (bs, 1H, C-4''-H), 5.15 (t, $J=7$ Hz, C-2''-H), 6.62 (d, $J=9$ Hz, 1H, C-6'-H), 6.78 (d, $J=2$ Hz, 1H, C-6-H), 6.94 (d, $J=9$ Hz, 1H, C-5'-H), 7.14 (d, $J=2$ Hz, 1H, C-8-H), 7.68 (1H, s, C-2-H); eims

m/z (rel. int.) 478 (M^+ , 39), 436 (M^+ -42, 40), 394 (436-42, 70), 352 (394-42, 19), 337 (352-15, 75).

CROTALARIN [3].—The product from the fractions 46 to 60 of silica column eluted with C_6H_6 -MeOH (95:5) was subjected to preparative tlc using C_6H_6 -MeOH (94:6) as eluent followed by crystallization, ($CHCl_3$ /MeOH) yielded crotalarin [3] (60 mg) mp 281-285°; yield 0.0012%; uv λ max (MeOH) 260 (log ϵ , 5.6152), 320 (sh) (log ϵ , 4.8386); λ max (MeOH) ($AlCl_3$) 260 and 360 nm; ir ν max (KBr) 3200, 1620, 1560, 1450, 1305, 1260, 1220, 1160, 1120, 1080, 1060, 1015, 870, 840 cm^{-1} ; 1H nmr δ ($CDCl_3$ +DMSO- d_6) 1.32 (s, 6H, C-4", 5" C-(CH_3)₂), 1.74 (t, $J=7$ Hz, 2H, C-2"- H_2), 2.6 (t, $J=7$ Hz, 2H, C-3", CH_2), 6.36 (s, 1H, C-6, CH), 6.75 (d, $J=9$ Hz, 2H, C-3', 5'- H) 7.32 (d, $J=9$ Hz, 2H, C-2', 6'- H), 7.62 (s, 1H, C-2- H); ^{13}C nmr refer to Table 1; eims m/z (rel. int.) 338 (M^+ , 100), 323 (M^+ - CH_3 , 17), 295 (M^+ -43, 51), 282 (M^+ -56, 28), 283 (M^+ -55, 100), 176 (25), 118 (21).

CROTALARIN DIACETATE [4].—A mixture of crotalarin 3 (50 mg), Ac_2O (0.5 ml), and pyridine (0.5 ml) was kept at ambient temperature for 20 h. The resulting mixture was worked up as for 2 to give the diacetate 4, mp 179-180° (MeOH); ir ν max (KBr) 1760, 1375, 1240, 1200, 1125, 1080, 1058, 920, 905, 875, 840 cm^{-1} ; 1H nmr δ ($CDCl_3$) 1.38 (s, 6H, C-4", 5"-C(CH_3)₂), 1.78 (t, $J=7$ Hz, 2H, C-2"- H_2), 2.22 (s, 3H, OAc), 2.3 (s, 3H, OAc), 2.58 (t, $J=7$ Hz, 2H, C-3"- H_2), 6.66 (s, 1H, C-6-H), 7.05 (d, $J=9$ Hz, 2H, C-3', 5'- H), 7.45 (d, $J=9$ Hz, 2H, C-2', 6'- H), 7.7 (s, 1H, C₂- H); eims m/z (rel. int.) 422 (M^+ , 16.2), 380 (M^+ -42, 29), 338 (380-42, 11), 295 (12), 283 (36).

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